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Blood Alcohol Content Analysis using **Nitrogen Carrier Gas**

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Blood Alcohol Content Analysis Using Nitrogen Carrier Gas

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Alcohol consumption can seriously affect the ability of a driver to operate a vehicle and blood alcohol content (BAC) directly correlates with this impairment. A number of nations have zero alcohol tolerance for motorists, but the majority of countries worldwide have a limit of between 50 and 80 mg alcohol per 100 ml blood, or 0.05-0.08%. Results are used in court to provide quantitative levels of BAC, which makes it one of the most commonly practised analyses in forensic laboratories. The large number of samples and requirement for speed of sample processing mean that analysis needs to be conducted quickly, whilst giving reliable and accurate results.

For analysis of BAC, headspace GC with FID detection is typically used. Headspace GC allows the quantitative analysis of alcohol directly from blood samples. Standard headspace systems use nitrogen for vial pressurization, with helium typically used for GC carrier gas. This application note looks at the use of nitrogen for both vial pressurisation and GC carrier gas. Nitrogen offers a cost-effective, abundant alternative to helium for carrier gas, whilst giving similar performance. Here we compare analysis of real forensic blood samples, taken from motorists suspected of driving under the influence of alcohol, analysed using nitrogen and helium carrier gas.

Sample Preparation

Using a Hamilton Microlab 600 Diluter, 200 μL of calibrators, controls, or blood samples were aliquoted and dispensed with 2000 μL of internal standard solution into a 10ml headspace

vial and capped. The internal solution consisted of 0.03% (v/v) n-propanol/ 1M ammonium sulfate/ 0.1 M sodium hydrosulfite. NIST traceable aqueous ethanol solutions from Cerilliant and Lipomed were used as calibrators (10, 50, 80, 200, 300, 500 mg/dL) and controls (20, 80, 400 mg/dL) respectively.

Experimental

Analyses were conducted using an Agilent 7890B GC with split/splitless inlet and dual columns each connected to an FID detector. Splitting of the samples onto the columns was via an Agilent unpurged Capillary Flow Technology splitter. The GC was coupled with an Agilent 7697A headspace sampler. Vial pressurization gas for all tests was provided by a Peak Scientific Precision Nitrogen Generator. Carrier gas was provided by either helium cylinder or the Precision Nitrogen Standard Generator. The HS-GC-FID system operating conditions are displayed in Table 1.

Headspace sampler	Agilent 7697A	
Vial pressurization gas	Nitrogen	
Oven Temperature	70	
Loop Temperature	70	
Transfer line	Deactivated fused silica, 0.53 mm id	
Transfer line temperature	90	
Gas Chromatograph	Agilent 7890B	
Carrier gas	Helium	Nitrogen
Detector	FID	
Columns	DB-ALC1 (30m x 320 μm x 1.8 μm), DB-ALC2 (30m x 320 μm x 1.8 μm)	
Split ratio	10:1	
GC Oven Start temperature	40°C (3 mins)	
GC Oven program rate	40°C min ⁻¹	
GC oven final temperature	120°C (1.2)	
Method runtime	6.2 minutes	

Table 1. HS-GC-FID Operating Conditions

The software used for analysis was Agilent MassHunter GC/MS Acquisition and MSD ChemStation Enhanced Data Analysis E.02.02.1431.

Results

Calibration curves produced with helium and nitrogen carrier gas both gave very good linearity with both curves having R² values of 99.9999.

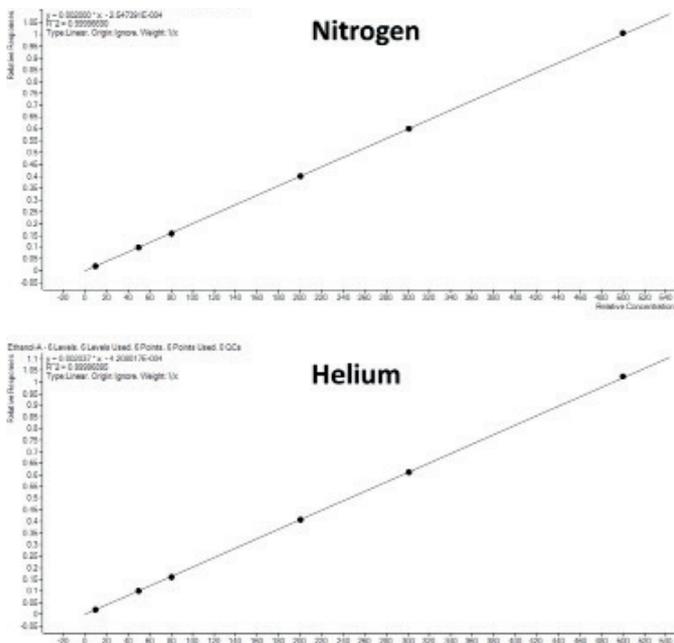


Figure 1: Calibration curves for ethanol standards run using nitrogen and helium carrier gas.

Blood alcohol levels of 5 blood samples were analysed. Figures 2 and 3 show chromatograms from the DB-ALC1 and DB-ALC2 columns, respectively for the separation and elution order of analytes for the multi-component resolution mix when run using nitrogen and helium carrier gas. Separation of potentially interfering components, such as methanol and 2-propanol was achieved within 3 minutes when using either carrier gas (Figure 2 & Figure 3).

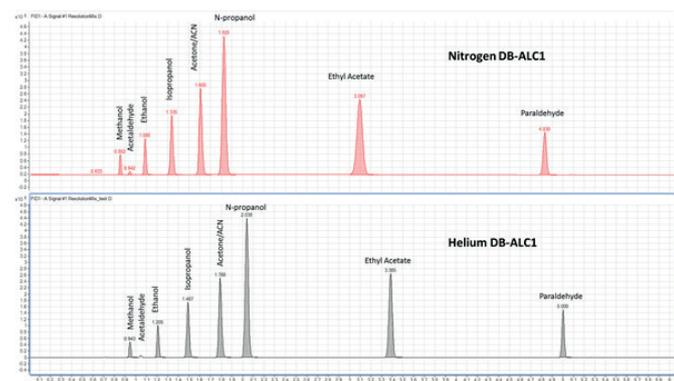


Figure 2: Results of resolution mixture run on DB-ALC1 column using nitrogen and helium carrier gas.

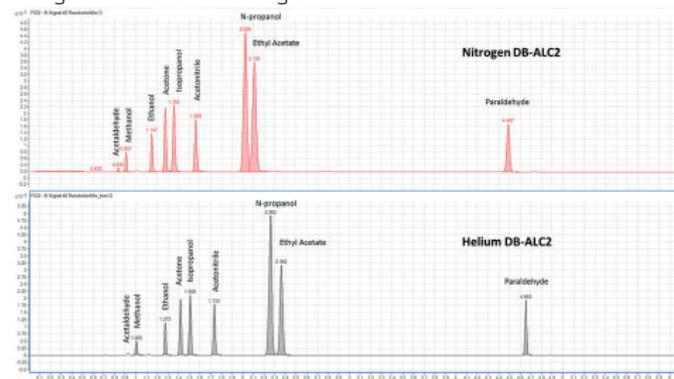


Figure 3: Results of resolution mixture run on DB-ALC2 column using nitrogen and helium carrier gas.

Results of analyses of real blood samples (analysed in duplicate) run with both nitrogen and helium carrier gas gave equivalent results with no differences found in the calculated ethanol concentrations (Table 2).

	Amount of ethanol detected (%)	
	Nitrogen	Helium
Sample 1A	0.05749	0.05702
Sample 1B	0.05776	0.05689
Sample 2A	0.01438	0.01421
Sample 2B	0.01433	0.01417
Sample 3A	0.23587	0.23476
Sample 3B	0.23481	0.23323
Sample 4A	0.02295	0.02254
Sample 4B	0.02285	0.02255
Sample 5A	0.05890	0.05866
Sample 5B	0.05948	0.05867

Table 2: Blood alcohol analysis results from analysis conducted with nitrogen and helium carrier gas.

Of the five blood samples tested, one was over 0.2%, which would result in a driving ban in almost every country worldwide. Two samples were over 0.05% which would result in a driving ban in a number of countries. The other two samples were 0.014% and 0.023% which would be below the limit in the majority of countries worldwide.

Conclusions

Results of BAC analysis show that there is no difference in the linearity of the calibration curve, or of the calculated ethanol content of real blood samples regardless of whether nitrogen or helium carrier gas was used. As an abundant, inexpensive alternative to helium, which is becoming increasingly more costly, there is no reason why nitrogen cannot be used for BAC analysis in place of helium. Since nitrogen is often used for vial pressurisation in headspace samplers, the use of a single gas source for vial pressurisation, carrier gas and FID make-up gas simplifies the lab's gas sourcing and would allow total gas supply from gas generators if the precision nitrogen was used in conjunction with the Precision hydrogen and zero air generators for GC-FID analysis.

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Greg Dooley Ph.D is the Director of the Analytical Toxicology Laboratory at Colorado State University. His research involves the development of analytical methods utilizing state of the art instrumentation to measure analytes of interest for current forensic and analytical toxicology applications.